



## Raman Spectra Evaluation of Gold Nanoparticles Composite Interaction with Nerve Agent Simulant

Siti Zulaikha Ngah Demon<sup>1,\*</sup>, Nurul Farhana Abu Kasim<sup>2</sup>, Norhana Abdul Halim<sup>1</sup>, Fadhlina Che Ros<sup>1</sup>, Nursaadah Ahmad Poad<sup>2</sup>, Nurul Syahirah Nasuha Saa'ya<sup>2</sup>, Ong Keat Khim<sup>1</sup>, Ngo Khoa Quang<sup>3</sup>

<sup>1</sup> Centre for Defence Foundation Studies, National Defence University of Malaysia, 57000 Kuala Lumpur, Malaysia.

<sup>2</sup> Faculty for Defence Science and Technology, National Defence University of Malaysia, 57000 Kuala Lumpur, Malaysia.

<sup>3</sup> University of Sciences, Hue University, 77 Nguyen Hue, Hue City, Vietnam

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### ABSTRACT

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Raman spectroscopy is one of many reliable methods to detect chemicals' presence and traces in environment. In present study, we evaluate Raman detection towards dimethyl methyl phosphate (DMMP), a colorless organophosphate chemical on selected substrates and environment. The role of AuNPs in providing surface-enhanced Raman scattering signals and graphene oxide as interaction template are discussed in the interest of analysing various DMMP contaminations. Our study shows that DMMP's Raman modes were enhanced by gold nanoparticles (AuNPs) coated cellulose film indicating potential of this substrate. In aqueous measurement, the D band to G band intensity ratio of AuNP/graphene was seen to produce significant increase in sensitivity towards DMMP compared to graphene oxide dispersion. These Raman signatures can be used to understand molecules-substrate's surface interactions that will be beneficial to chemical and optical sensors development. The current system also shown the potential to identify 1000 ppm DMMP which is difficult by typical infrared spectroscopy.

## 1. Introduction

Exposure and misuse of organophosphate compounds originated from insecticides, drugs and chemical warfare agents (CWA) are potential hazard to health and environment [1]. CWA organophosphate nerve agents are highly toxic capable to accelerate acute symptoms to death within minutes of poisoning. Chemical detection is one of the four counter strategies (deter, detect, delay, and defend) used by national securities to protect vulnerable against this threat typically by employing various sensing instruments ranging from electrical, optical, radioactive, electrochemical, mass spectrometric and other possible methods [2]. As optical method, Raman spectroscopy is useful as complementary to infrared spectroscopy in which the later probes molecular absorption at specific vibrational frequencies that causes a change in the dipole moment while Raman relies on the change in the polarizability of a molecule at the Raman shift frequencies at which the molecule scatters radiation. Though typical cross-section of Raman scattering is only  $\sim 10^{-29}$  cm<sup>2</sup> per molecule, signal

\* Corresponding author.

E-mail address: zulaikha@upnm.edu.my. (S.Z.N. Demon)

enhancement by means of electronic resonance and electromagnetic resonator allow better sensitivity in detection as far as a single molecule detection [3-5].

Surface enhanced Raman spectroscopy (SERS) was first reported by Fleischmann and co-workers in 1974 whose intense Raman signal arised from pyridine adsorption on Ag electrodes. After that, Raman application in surface adsorption studies rapidly grew contributed to surface plasmons excitations mainly observed in Ag, Au and Cu based structures [3]. Signal enhancement was also reported by other metals such as Pt, Fe, Co and Ni, however, on a lesser scale. Gold (AuNP) and silver (AgNP) nanoparticles are known as feasible substrates to detect dimethyl methyl phosphate (DMMP), an organophosphate known as simulant to Sarin [7-8]. These substrates are believed to provide high electromagnetic enhancement and chemical affinity towards target molecule while avoiding band interference with responsible functional groups [4,7]. Mode selection by SERS substrates provides additional information on molecule symmetry when adsorbed to a surface which differs when it is in freeform [9].

Metal incorporated graphenous materials such as graphene oxide (GO) and reduced graphene oxide (rGO) are employed as optical based biosensors due to signal enhancement by charge transfer from porous graphene to metal [5,10-12]. Metal alone has poor adsorption of biomolecules, but with graphene addition as composite, adsorption efficiency increases and creates stronger electric field at the interface [10]. An investigation by Rattan and his co-workers has shown that AuNPs functioned as electron mediator for the transfer of electrons from rGO to NO<sub>2</sub> gas as a highly efficient gas sensor [13]. High surface area of graphene and GO-based materials facilitate adsorption of target molecules, and their sensing capability can be derived by favorability of  $\pi$ - $\pi$  interactions occurring within the aromatic rings [14]. The same graphene surface plays a key role in making AuNPs to be efficiently integrated and decorated, thus, its influence in optical detection will be highly interesting as well.

Known for its practicability, handheld Raman device is suitable for in-situ/real-time identification of dangerous chemical residues in solid, liquids and gaseous forms. In this paper, feasibility of the system to identify DMMP using several AuNPs coated surface as SERS substrate has been demonstrated. Similar techniques were also employed to investigate role of metal nanoparticles in improving analyte adsorption by graphenes. Our evaluation of Raman spectra can provide better understanding of AuNP and graphene surface interactions at both ambient and aqueous conditions and also provide recommendation in utilizing nanomaterials for chemical detection.

## **2. Methodology**

### *2.1 Sample preparation*

Graphene oxide (GO dispersion in H<sub>2</sub>O, 4 mg/ml), gold (III) chloride hydrate (HAuCl<sub>4</sub>, 99.995% trace metal basis), sodium citrate and ascorbic acid were purchased from Sigma Aldrich. Deionized water was prepared by a Milli-Q Plus system (Millipore). GO aqueous dispersion was prepared by 30 min sonication of 1:30 of GO:deionized water. Gold nanoparticles (AuNPs) were reduced from aqueous HAuCl<sub>4</sub> using sodium citrate. One pot reduction of AuNP-graphene was prepared according to our previous investigation [15]. Briefly, 100 ml of GO dispersion and 20 ml of 0.01 M HAuCl<sub>4</sub> was mixed and stirred before adding 30 mg of ascorbic acid. After that, the solution was heated to 95°C and kept for 90 min under vigorous stirring, resulting into AuNP/graphene composite.

The substrates were carefully prepared to provide homogenous AuNPs deposition. Silane treated quartz substrate in ref. 15 was dip coated in aqueous AuNPs for 24 h to produce AuNPs coated glass while AuNPs coated Whatman paper was collected by filtering the same aqueous AuNPs using conical glass funnel and Whatman paper No.1. The substrates were subsequently left to dry for 48 h. For the AuNPs cellulose film, cellulose acetate powder was stirred in acetone at ambient for 6 h prior to

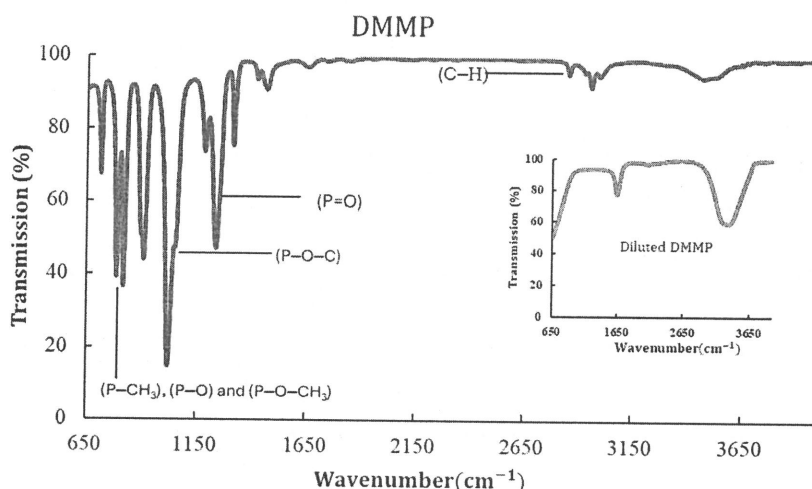
addition of the aqueous AuNPs. The mixture continued to stir until homogenous before it was poured onto glass petri dishes and left to dry. The films can be peeled away when the solvent completely dried.

## 2.2 FTIR and Raman measurements

FTIR study was carried out with attenuated total reflectance accessory configuration in the mid IR range provided by Perkin Elmer Frontier IR instrument. Resolve Handheld Raman Analyzer by Agilent Technologies was used to obtain Raman spectra. The laser is 830 nm with maximum irradiance of 475 mW. All spectra were collected in  $350\text{ cm}^{-1}$  –  $2000\text{ cm}^{-1}$  spectral range,  $1\text{ cm}^{-1}$  spectral resolution and in back-scattering geometry. The technology provided by the Raman system allows chemical identification through its container. Dimethyl methyl phosphonate ( $\text{CH}_3\text{P}(\text{O})(\text{OCH}_3)_2$ ,  $M_w=124.8\text{ g mol}^{-1}$ ) was purchased from Sigma-Aldrich. The analyte was used without purification. DMMP of 1000 ppm, 5000 ppm, 10 000 ppm, 20 000 ppm, 50 000 ppm and 70 000 ppm dilutions were made with deionized water. These dilutions were lightly shaken and dropped into sample or substrates before Raman measurement was taken. The distance between the Raman nose and the substrates was approximately 1 cm apart.

## 3. Results and Discussion

### 3.1 Identification of DMMP and diluted DMMP using FTIR and Raman



**Fig. 1.** ATR-FTIR spectrum of undiluted DMMP solution and 5000 ppm DMMP dilution (subset). Water dominated infrared absorption peaks which interfered identification.

Figure 1 shows FTIR spectra obtained after pure DMMP and diluted DMMP solution were dropped onto ATR surface. This condensed phase of DMMP consistently agreed with ref. 16 with strongest absorption observed at  $1050.1\text{ cm}^{-1}$  for asymmetric stretching of P-O-C, P=O stretching mode located at  $1274\text{ cm}^{-1}$ , and lastly, C-H stretching modes appearing around  $2800\text{--}3100\text{ cm}^{-1}$ . Also presented in the figure is compilation of P-CH<sub>3</sub>, P-O and P-O-CH<sub>3</sub> stretching/bending modes around  $700\text{--}800\text{ cm}^{-1}$ , the fingerprint region. The subset of Figure 1 shows spectra of 1000 ppm and 5000 ppm diluted DMMP which presents none of these strong features, indistinguishable from typical ATR-

FTIR spectra measured from water. Upon closer inspection of the spectra, we recognized ~0.5 % absorption change at similar vibration regions for DMMP. Raman spectra of DMMP shows a strong peak belonging to the phosphate group vibrational mode located at 715 cm<sup>-1</sup> for both undiluted and diluted DMMP (Fig.2). This peak can be observed in spectrum of 5000 ppm DMMP without strong interference by water. This unique feature in Raman suppressed the strongest bond observed in FTIR and provided complementary investigation of the functional group analysis together with IR spectroscopy.

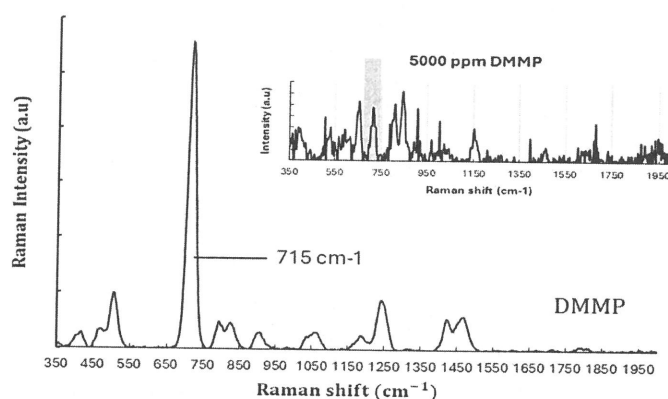


Fig. 2. Raman spectra of undiluted DMMP and 5000 ppm DMMP dilution (subset).

### 3.2 Raman quantification of DMMP

First of all, the quantification analysis has to be established for the Raman system. A calibration plot as shown in Fig. 3 was produced using two different DMMP dilutions. Raman peak of 715 cm<sup>-1</sup> previously identified as DMMP's has increased by 5 times with 5000 DMMP dilution. Lafuente et al has established this peak as monitoring peak in their Raman study [7]. This linear correlation implied that the current method assessment is suitable for quantification purpose. Bear in mind, there are several other environmental factors that should be accounted for when taking the measurement such as the distance between the laser nose cone and the sample, type and thickness of substrates and others. These factors are consistently controlled throughout the experiments.

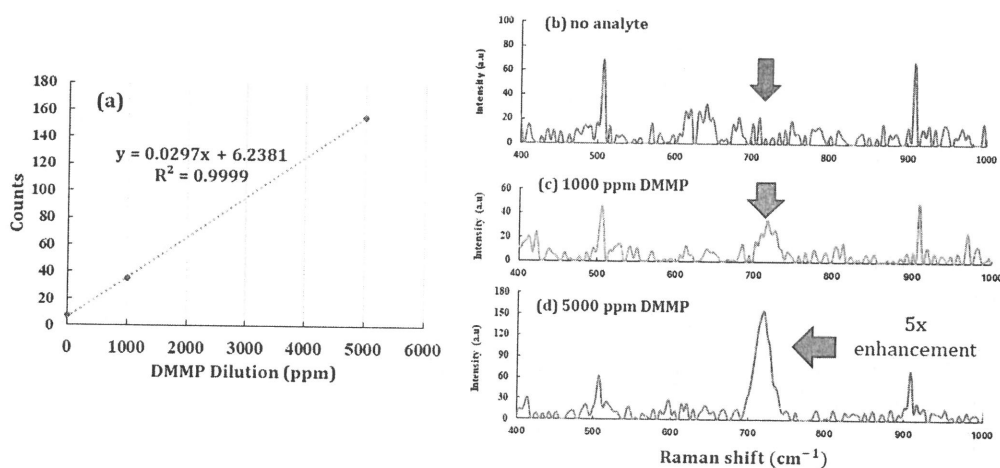


Fig. 3. (a) Calibration plot of DMMP dilutions and Raman spectra of (b) clean substrate, (c) 1000 ppm DMMP and (d) 5000 ppm DMMP.

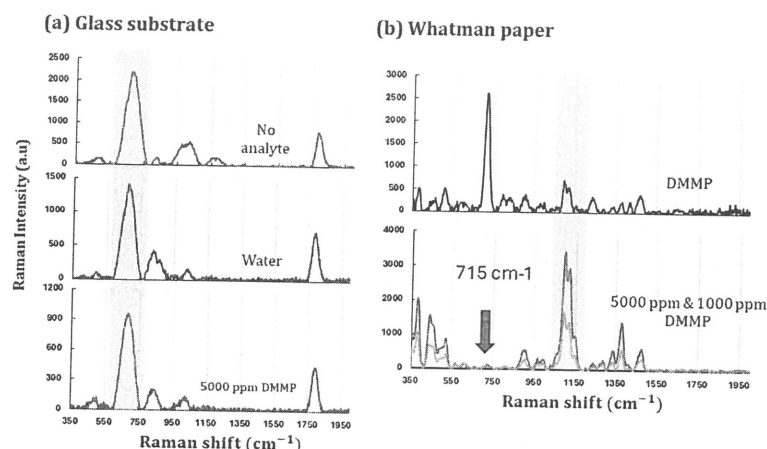


Fig. 4. Some Raman spectra of analytes collected from (a) glass substrate and (b) Whatman paper.

Interference from Raman signal is possible with certain choice of substrates in addition to low concentration or weak signal contribution from the compound of interest. Two common substrates found in the laboratory were used to demonstrate substrate interference in Raman identification. In the case of DMMP assessment in Fig. 4, we found the broad Raman signal from glass substrate centred at 700 cm<sup>-1</sup> has overlapped with the most prominent Raman peak of DMMP. The presence of dominant peak from other regions can also affected the sensitivity from analytes such as shown in Raman spectra obtained from Whatman paper. Large contribution from C-O vibrational modes in some way has added the difficulty to distinguish the 715 cm<sup>-1</sup> peak from the spectra of 5000 ppm DMMP dilution. Thus, there is a need to employ Raman enhancement strategies when obtaining both qualitative and quantitative analysis of analyte.

### 3.3 Raman of DMMP from AuNPs Cellulose Film

Cellulose film is solution for flexible and free standing substrate, hence, is proposed as SERS substrate. In this investigation, AuNPs cellulose film is reproduced from Khalaf et al [17]. Figure 5 shows the UV-Vis absorption spectrum of the AuNPs cellulose film which enclosed both absorption peaks of cellulose around 270 nm and SPR peak of AuNPs centered around 580 nm. Since the SPR does not neither coincides nor is close to the energy of incident photon, we can neglect the possibility of resonant Raman scattering in our method [18-19].

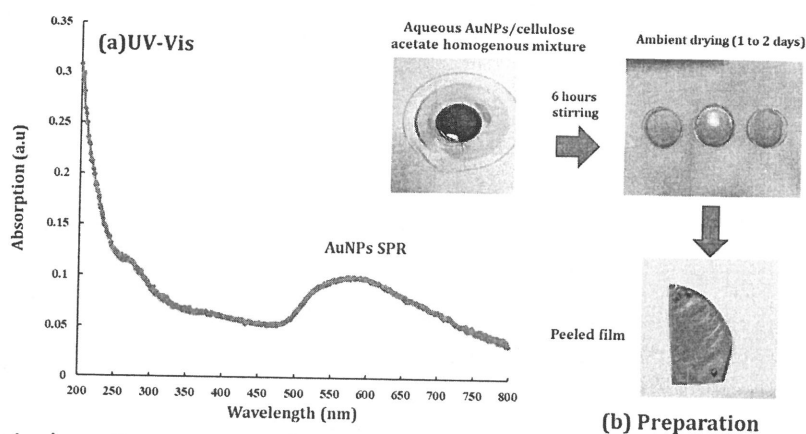


Fig. 5. (a) UV-vis absorption spectrum of AuNPs cellulose film. (b) Preparation steps to produce the film.

Raman spectra collected from AuNPs coated cellulose film before and after 5000 ppm DMMP dilution dropped onto the surface are presented as Fig. 6 (a). No substantial signal was observed when using pristine cellulose surface as the analyte quickly adsorb onto the film layer. Apparently, AuNPs content has reduced the hydrophilicity of cellulose which increases its reusability with liquid analyte for several times. In addition, several Raman peaks appeared besides the prominent  $715\text{ cm}^{-1}$  peak where their origin can be traced back to the pure DMMP spectrum seen in Fig. 2. These moderately strong peaks manifested in cellulose-based film due to clean background interference from the substrates. Shift of positions of DMMP's fingerprint peaks can be analysed further to investigate molecular geometry of DMMP-Au complex formed during interaction [7].

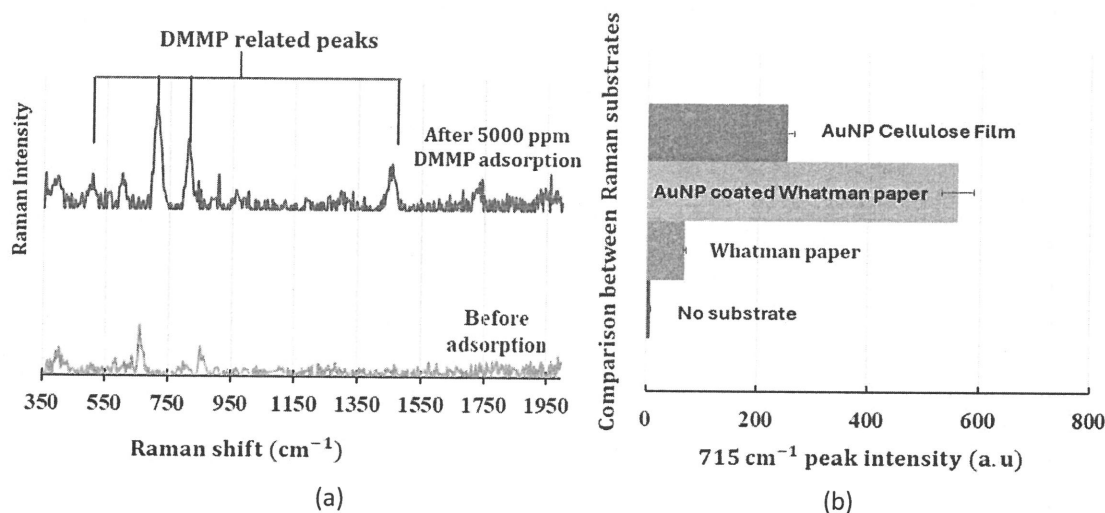


Fig. 6. (a) Raman spectra obtained before and after 5000 ppm DMMP was dropped onto the AuNPs cellulose film. (b) Comparison of DMMP's Raman peak from different substrates tested in this study.

Table 1 summarize Raman spectra evaluation when 5000 ppm of DMMP analyte were introduced to surface of different substrates. Dominant Raman signal from glass is seen within the same range of DMMP should appeared for AuNPs coated glass sample, while the  $715\text{ cm}^{-1}$  peak of DMMP materialized more clearly for AuNPs coated Whatman paper compared to the uncoated one. This Raman enhancement by the AuNPs coated Whatman paper is visualised in Fig. 6 (b) where its intensity is around 9 times of Raman intensity obtained from the uncoated paper. Nevertheless, this enhancement ratio is much lower than typically achieved by SERS perhaps due to technical limitation in handheld system and rough surface of Whatman paper that produced many scattering centres. Inefficient detection from AuNPs coated glass substrate is because of the insufficient film coverage on the glass surface, thus can be improved by exploring other deposition methods. We also explored the possibility of insufficient AuNPs aggregation formation that should have manifested as dark red color colloid, thus the present AuNPs can pose inadequate material for SERS study [18].

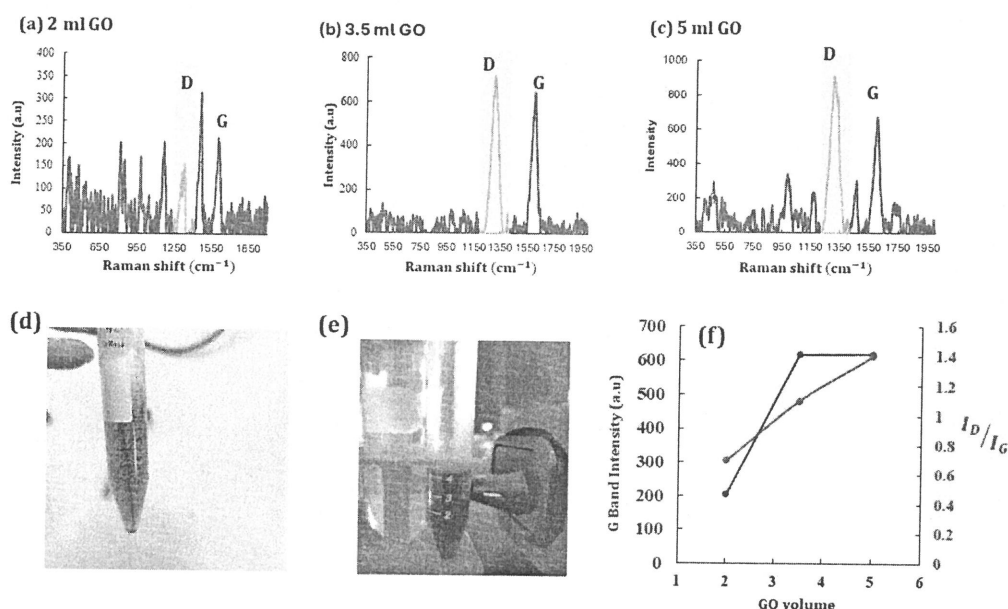
**Table 1**  
Observation from different Raman substrates for DMMP identification

Substrate	Observation
AuNPs coated glass	Dominant Raman contribution from glass.
AuNPs coated Whatman paper	Strong $715\text{ cm}^{-1}$ peak was observed.
AuNPs Cellulose film	Several fingerprint peaks belonging to DMMP appeared.

### 3.4 Raman of DMMP from Graphene- and AuNP/Graphene in Aqueous Environment

Raman of graphene is one of fundamental data indicating quality of graphene in term of structure, conductivity and presence of dopants [20]. Intensity ratio,  $I_D/I_G$  measured from the intensity of defect, D band ( $\sim 1350\text{ cm}^{-1}$ ) and graphene, G band ( $\sim 1580\text{ cm}^{-1}$ ) are the basis of the degree of inter-defect distance in graphene [20-21]. G band is related to vibrations of  $sp^2$  domains and several defect-associated bands while D band arises when there are defective sites in the C aromatic lattice [21]. Previous research has established graphene's role as substrate, building block and also Raman probe in hybrid combination with metal nanoparticles [6]. At ambient, surface of graphene is weakly interacting surface, therefore chemical sensing can only be derived from its functionalization [14]. Surface interaction between graphene and DMMP was done in aqueous environment where the hydrophobic nature with dispersant must be considered, and their interaction will be evaluated using G and D bands of graphene. We believed that this environment gives valuable insights towards future making of SERS substrate.

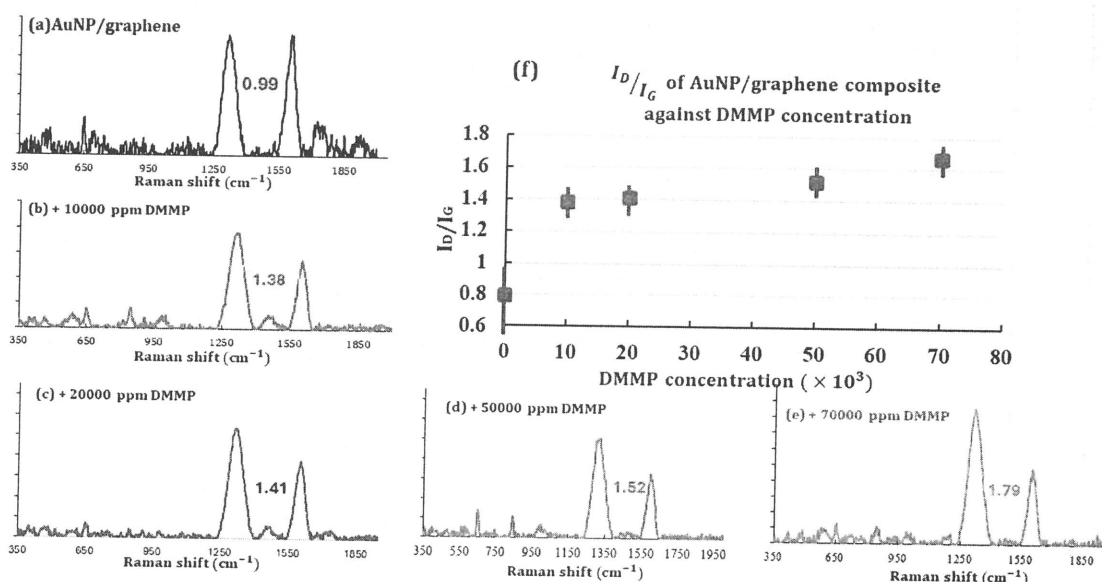
Figure 7 (a-c) show Raman spectra obtained from different GO volumes mixed with 2 ml pure DMMP analytes. The dispersed GO was contained in plastic test tube (Fig. 7(d)) and positioned in front of nose cone of the Raman spectrometer as shown in Fig. 7 (e). Large background noise was detected in the spectra, fortunately, the D and G bands of the GO becomes more prominent with increasing GO volumes. This linear proportionality is illustrated in Fig. 7 (f) using G band intensity because of its position and overlapping intensity that is less susceptible to any defects on graphene planes.



**Fig. 7.** Raman spectra from GO colloid samples (a) 2 ml GO + DMMP, (b) 3.5 ml + DMMP and (c) 5 ml + DMMP. (d) Sample plastic tube containing mixture of GO and DMMP and (e) the sample is positioned directly to the nose cone of the Raman system. (f) Relationship between graphene band intensities with the GO volume.

On the other hand, the D band intensities change drastically in ratio to the G band with increasing GO volume. Increasing  $I_D/I_G$  is often associated with increase of carbon to oxygen ratio, and also expanding interlayer distance between graphene basal planes [21]. The change from 0.7 to 1.4 can be related graphene surface preferential toward DMMP molecules over water molecules. The 2 ml

GO possessed less surface area for DMMP occupation compared to 3.5 ml and 5 ml GO. Large amount of DMMP molecules on graphene surface may also affect surface charge which leads to drastic increase of  $I_D$ . Interestingly, we did not obtain similar result when the same experiment was repeated with 5000 ppm DMMP dilution which indicated insensitivity detection by GO.



**Fig. 8.** Raman spectra of (a) AuNP/graphene composite and (b-e) the composite after mixed with DMMP dilutions. Figure (f) is plot of  $I_D/I_G$  ratio in Raman spectra from AuNP/graphene aqueous samples as a function of different concentration of DMMP dilutions.

Figure 8 shows the spectra obtained by AuNP/graphene composite that implied remarkable sensitivity toward DMMP. It was seen that the  $I_D/I_G$  ratio increase from 0.99 to 1.79 as the higher concentration of DMMP is added subsequently onto the same sample. This different turn out when AuNP/graphene was used to substitute the GO means that AuNP incorporation is a crucial factor in enhancing the sensitivity of DMMP adsorption on graphene surfaces. Firstly, relatively high ratio by AuNP/graphene in Fig. 8 (a) indicating nanoparticles was successful anchored on graphene edges [15, 22]. The presence of AuNPs within the graphene planes posed weaker van der Waals interaction as observed from formation of crumpled graphene sheets in HRTEM image from ref. 15. This graphene structural disorder induced larger interlayer distance in graphene stacks compared to GO. The already weakly interacting graphenes will disperse further after more DMMP molecules is added into the composite. The preferential of DMMP to be adsorbed on the aromatic rings will cause increase of D band intensity as seen in Fig. 8 (b-e). In summary, the increase of D band intensities could be directly related to increase of graphene interlayer spacing that reduce bulk aggregation of rGO. AuNPs doping assisted re-aggregation mechanism even at low DMMP concentration. This microscopic explanation can be used to produce selectivity of material against different analytes which subject to their polarity and hydrogen bonding interactions.

The fact that the D band to G band intensity ratio of AuNPs/graphene show remarkable sensitivity towards DMMP dilutions compared to GO indicated that the former can provides better sensing surface in optical detection. The non-linear relationship shown in the graph also implied that the variation between  $I_D/I_G$  ratio and the DMMP concentration may reach point of saturation after certain addition, hence might require more optimization for this material strategy. Under these

circumstances, no sensing event related to charge transfer from Au to graphene is detected. Charge transfer between molecules should manifest in amplification of Raman signals belonging to DMMP. Another challenge in achieving SERS signal in the form of one-pot reduction of AuNP/graphene is the inhomogeneous accumulation of particles on the edges where there is higher presence of oxygen functional groups (-OH and -COOH) compared to basal planes [5].

Lastly, the reliability of this evaluation is presented in the form of average relative standard deviation of  $I_D/I_G$  at 5% as the measurement from aqueous sample were taken repetitively over 8 minutes. The result is shown in Fig. 9. This test shows that the AuNP/graphene composite remained stable even after several laser radiation. The reliability and stability of both measurement technique and the sensing material are crucial in detection of nerve agent chemicals and as demonstrated from this study, both components have the potential to be explored in defence and security applications.

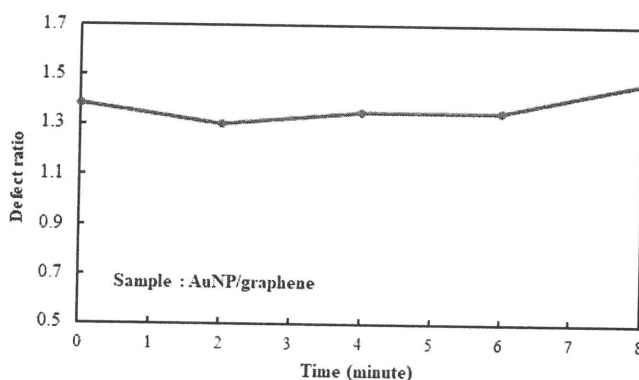


Fig. 9. Stability test for Raman measurement taken from AuNP/graphene samples in aqueous environment.

#### 4. Conclusions

The interaction between gold nanocomposites and DMMP in various forms was studied using Raman spectra. The Raman system is not a conventional method to yield quantitative information, but we evaluate its qualitative reliability with SERS method. Based on this result, we highlight some vibrational modes that are crucial in organophosphate identification. Issue of overlapping vibrational modes due to interference from substrates may hindered Raman measurement, nevertheless it is believed that by optimizing graphene and gold nanoparticles sizes, the method can deliver more accurate analysis in chemical trace identification. DMMP identification in AuNP/graphene composite in aqueous environment yield promising result in probing molecules adsorption on graphene surface due to preferential interaction in analytes.

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